# Presence of cell-free fetal circRNA in maternal plasma

Yuxin Ran<sup>1,2,3</sup>, Ruixin Chen<sup>4</sup>, Yangyu Zhao<sup>5</sup>, Nanlin Yin<sup>6</sup>, Hongbo Qi<sup>1,7</sup>

- <sup>1</sup>Department of Obstetrics, The First Afliated Hospital of Chongqing Medical University, Chongqing, China
- <sup>2</sup>Chongqing Key Laboratory of Maternal and Fetal Medicine, Chongqing Medical University, Chongqing, China
- <sup>3</sup>Joint International Research Laboratory of Reproduction and Development of Chinese Ministry of Education, Chongqing Medical University, Chongqing, China,
- <sup>4</sup>Department of Gynecology and Obstetrics, West China Second Hospital, Sichuan University, Chengdu, China
- <sup>5</sup>Department of Obstetrics and Gynecology, Peking University Third Hospital, Beijing, China
- <sup>6</sup>Center for Reproductive Medicine, The First Affiliated Hospital of Chongqing Medical University, Chongqing, China
- <sup>7</sup>Chongqing Health Center for Women and Children, Chongqing, China

Submitted: 28 November 2022; Accepted: 28 January 2022 Online publication: 23 February 2022

Arch Med Sci 2022; 18 (2): 540–544 DOI: https://doi.org/10.5114/aoms/146204 Copyright © 2022 Termedia & Banach

#### Abstract

**Introduction:** There is an urgent need to find novel stable cell-free fetal (cff-) RNA in the maternal circulation to facilitate the advance of non-invasive prenatal testing (NIPT) to more effectively avoid birth defects.

**Methods:** CircRNA microarray was used to detect the cff-circRNA in plasma. **Results:** There were cff-circRNAs from the fetus in the peripheral blood of pregnant women and they persisted even until at least 24 h after delivery. In addition, we found that cff-circRNA might have a specific expression pattern in gestational disease.

**Conclusions:** We demonstrated the presence of cff-circRNA in the maternal circulation, which may shed new light on the development of NIPT.

**Key words:** noninvasive prenatal testing, cell-free fetal circRNA, maternal-fetal communication.

In 1997, the presence of cell-free fetal (cff-) DNA in maternal plasma was demonstrated, ushering in a new era of noninvasive prenatal testing (NIPT) [1]. Over the past two decades, this burgeoning revolutionary strategy has considerably improved the early detection of birth defects worldwide [2]. Currently, cff-mRNA is emerging as a promising biomarker for NIPT beyond cff-DNA, as it can not only be used to screen for genetic defects such as aneuploidy but also be theoretically considered to have far more potential than DNA for real-time monitoring of fetal physiopathology [3]. However, cff-mRNA is susceptible to degradation and lacks stability and integrity as a marker, which greatly limits the feasibility of its clinical application [4]. Therefore, searching for a novel type of stable cff-RNA in the maternal circulation is particularly important for NIPT to break through the current dilemma. Here, for the first time, we report the presence of a stable RNA derived from fetal circulation, cff-circRNA, in maternal circulation, which might have implications for the NIPT strategy to monitor fetal biological alterations.

#### Corresponding authors:

Nanlin Yin Center for Reproductive Medicine The First Affiliated Hospital of Chongqing Medical University 1 Youyi Rd Yuzhong District Chongqing 400016, China E-mail: yinnanlin@cqmu. edu.cn

Hongbo Qi Chongqing Health Center for Women and Children 120 Longshan Road Yubei District Chongqing 401120, China E-mail: qihongbocy@gmail. com



Attribution-NonCommercial-ShareAlike 4.0 International (CC BY -NC -SA 4.0). License (http://creativecommons.org/licenses/by-nc-sa/4.0/).

Creative Commons licenses: This is an Open Access article distributed under the terms of the Creative Commons

Methods. Sample collection and processing. The pregnant women who came to the First Hospital of Chongging Medical University for a delivery hospitalization were recruited. All of them had singleton fetuses and were without serious pregnancy complications such as preeclampsia and fetal distress. With informed consent, their peripheral blood was collected in the third trimester (gestational week:  $34.6 \pm 4.0$ ) and after delivery using EDTA anticoagulant tubes. Then, the blood samples were centrifuged twice at 3000 rpm at 4°C to obtain plasma and stored at -80°C until testing [5]. Based on the pregnancy outcome, we finally selected samples from seven pregnant women carrying male fetuses for this study. The clinical information of these patients is shown in Supplementary Table SI.

The study was approved by the Medical Research Ethics Committee of the First Affiliated Hospital of Chongqing Medical University.

CircRNA microarray analysis. Total RNAs were extracted from plasma samples and purified using Trizol reagent (Invitrogen, Gaithersburg, MD, United States) and the NucleoSpin R RNA cleanup kit (MACHEREY-NAGEL, Germany), respectively. Then, the linear RNAs were removed by RNase R (Epicentre, Illumina, Inc.). The remaining RNAs were sequentially processed as follows: 1) reverse transcribed to first strand cDNA; 2) second strand cDNA synthesized; 3) cRNA synthesized using T7 Enzyme Mix; 4) reverse transcribed to cDNA; 5) reacted with Random Primer; 6) labeled with dNTP (Cy3-dCTP, Cy5-dCTP) with fluorescent moieties. The DNA with fluorophore was hybridized with the circRNA microarray in the hybridization mixture. Finally, the microarray was scanned using an Agilent microarray scanner (G2565CA); data were extracted using Agilent Feature Extraction (version = 10.7) software and normalized using Agilent GeneSpring software. All the processes of microarray analysis were conducted by the Bioassay Laboratory of CapitalBio Corporation (Beijing, China). The detailed information of experimental procedures is listed in Supplementary material.

**RNA-seq data collection and analysis.** We searched public databases for fetal/newborn peripheral blood sequencing datasets. Inclusion criteria were as follows: 1) the blood sample was collected immediately after birth for sequencing; 2) the information on the sex of the sample donors was provided; and 3) the data were in \*.fastq format. Ultimately, the RNA-seq raw data of peripheral blood from male and female newborns were obtained from the SRA (https://www.ncbi.nlm.nih.gov/sra) database (accession ID: SRP182878).

The fastq files were processed by the BWA-MEM algorithm to map the reads to the UCSC human reference genome (hg19). Then, the circRNAs were identified and quantified using the CIRI2 software, which works based on the recognition of the back-spliced junction reads. All the processes were used with the default parameters.

The count data of circRNA were normalized by the TMM algorithm of the R package "edgeR".

*Ethics.* The studies involving human participants were reviewed and approved by the Medical Research Ethics Committee of The First Affiliated Hospital of Chongqing Medical University (ID: 2020-60) and were subject to the Declaration of Helsinki. The patients/participants provided their written informed consent to participate in this study.

**Statistical analysis.** Statistical analysis was conducted using GraphPad Prism (version 8.0, San Diego, CA, United States). The relative expression levels of cff-circRNAs were represented by the total concentration of the circRNAs derived from the Y chromosome (Y-circRNA). The visualization of results was performed by GraphPad Prism and the R package "gglpot2".

Results. The detection of RNA and DNA derived from the Y chromosome of pregnant women with male fetuses demonstrated the presence of cff-RNA and cff-DNA in the maternal circulation, which led to the discovery of NIPT [1, 6]. With this, we speculate that if circRNA derived from the Y chromosome (Y-circRNA) can be found in the maternal circulation, it may further promote the development of NIPT. To confirm its feasibility, we analyzed circRNA expression profiles in peripheral blood of male and female neonates using RNA-seq data. The results showed that Y-circRNA was detectable in all the male neonates (n = 19), while it was negative in all the female samples (n = 12)(Supplementary Table SII). Thus, the presence of cff-circRNA can be proved by the detection of Y-circRNA in the maternal circulation.

Based on this, we examined the cff-circRNA in the circulation of 7 pregnant women with a male fetus before delivery via a microarray targeting Y-circRNA. Of the total 198 Y-circRNAs to be measured, 88 were detected in at least 3 samples, and 27 of them were detected in all 7 samples (Figures 1 A, B). This indicates that cff-circRNA does exist in the maternal circulation, and most of these detected Y-circRNAs are derived from the genes UTY, USP9Y, DDX3Y and ZFY, which is consistent with our findings in the peripheral blood of male newborns mentioned above (Figure 1 C).

Is the cff-circRNA in the maternal circulation sufficiently stable? In view of the instability of cff-mRNA in the circulation (the half-life is only 14 min), and the fact that it is quickly cleared after delivery [7], we performed the same test on the plasma of 7 pregnant women 24 h after delivery. The results showed that 66 of the 88 cff-circRNAs



Figure 1. Characterization of Y-circRNA in maternal circulation. Y-circRNA expression profiles in (A) prenatal and (D) postpartum maternal plasma. B – Statistics of the counts of Y-circRNAs detected in prenatal maternal plasma samples. C – The percentage of Y-circRNA of different genetic origins in the prenatal maternal circulation (left panel) and in the neonatal circulation (right panel). E – The intersection of Y-circRNAs in prenatal and postnatal maternal plasma

in prenatal plasma were still detectable in more than 3 postpartum plasma samples (Figures 1 D, E). This indicates that the existence of cff-circRNA (at least Y-circRNA) in the maternal circulation is stable, and may even last for a long time after delivery.

What is the significance of these circRNAs that are stably present in the maternal circulation? We preliminarily analyzed the association between cff-circRNA levels and clinical phenotypes to explore whether cff-circRNA may reflect underlying maternal-fetal biological alterations. We found that the relative expression levels of cff-circRNAs (represented by total concentrations of Y-circRNA) were higher in the prenatal plasma of pregnant women who eventually delivered prematurely than in that of full-term pregnancies (Figure 2). This suggested that more cff-circRNA may be transported from fetal to maternal circulation in the presence of pathological changes in the maternal-fetal system. That is, cff-circRNA might have potential as a biomarker for prenatal disease screening.

Discussion. In this study, we have demonstrated for the first time the presence of cff-circRNA in maternal plasma and found its potential association with diseases during pregnancy. This is a pioneering study and the results are preliminary, so large-scale clinical research is needed to validate and explore them in depth. A further limitation is related to the difficulties prevailing in current circRNA research. As a "young" RNA molecule, circRNA's recognition algorithm and naming system are far less mature than those of traditional RNAs, and there is even some confusion. This poses a challenge to our research. Nevertheless, our findings have good clinical application and promotion value. Firstly, circRNA is an emerging RNA with a unique closed-loop structure distinct from traditional linear RNAs (mRNA, miRNA, lncRNA) and thus has the property of stable existence against nucleases [8, 9]. So our findings have the potential to help solve one of the biggest shortcomings of the current clinical application of cff-RNA. Secondly, compared with traditional linear RNA, circRNA has more significant tissue and spatiotemporal specificity; thus, in theory, cff-circRNA may reflect fetal pathological alterations more accurately [10, 11]. Thirdly, current studies on circRNA in maternal blood do not take into account the presence of cff-circRNA, meaning that cff-circRNA is also counted and analyzed as maternal circRNA. This might introduce potential errors into the study results. So, it is valuable to draw the attention of researchers to this issue via our findings.

In conclusion, we have identified a new type of fetal nucleic acid in the maternal circulation that may shed new light on the development of



Figure 2. Relative level of cff-circRNA in prenatal maternal plasma of preterm and full-term pregnancies.

NIPT techniques and influence the achievement of real-time monitoring of fetal pathological alterations. It deserves to be explored by more systematic and extensive studies.

## Acknowledgments

Yuxin Ran and Ruixin Chen contributed equally to this work and should be considered co-first authors.

This work was supported by grants from the Chongqing Municipal Health Commission (No. 2020MSXM029), The Chongqing Municipal Education Commission (No. CYB20141), and The Science and Technology Department of Sichuan Province [No. 2020YFQ0006].

The authors thank E Gong of the Department of Obstetrics, the First Affiliated Hospital of Chongqing Medical University, for administrative support.

The authors would like to appreciate the support from "111 program" of Ministry of Education P.R.C and State Administration of Foreign Experts Affairs P.R.C.

### **Conflict of interest**

The authors declare no conflict of interest.

#### References

- 1. Lo YM, Corbetta N, Chamberlain PF, et al. Presence of fetal DNA in maternal plasma and serum. Lancet 1997; 350: 485-7.
- 2. Skrzypek H, Hui L. Noninvasive prenatal testing for fetal aneuploidy and single gene disorders. Best Pract Res Clin Obstet Gynaecol 2017; 42: 26-38.
- 3. Tamminga S, van Maarle M, Henneman L, et al. Maternal plasma DNA and RNA sequencing for prenatal testing. Adv Clin Chem 2016; 74: 63-102.
- 4. Mader S, Pantel K. Liquid biopsy: current status and future perspectives. Oncol Res Treat 2017; 40: 404-8.
- Alomari MA, Al-sheyab NA, Shattnawi KK, Khabour OF. Gender-specific differences in plasma ferritin in adolescents smoking cigarettes versus waterpipe smoking: the Irbid-TRY Project. Arch Med Sci 2021; DOI: https:// doi.org/10.5114/aoms/115011.
- 6. Poon LL, Leung TN, Lau TK, Lo YM. Presence of fetal RNA in maternal plasma. Clin Chem 2000; 46: 1832-4.

- 7. Chiu RW, Lui WB, Cheung MC, et al. Time profile of appearance and disappearance of circulating placentaderived mRNA in maternal plasma. Clin Chem 2006; 52: 313-6.
- 8. Huang Y, Zhang C, Xiong J, Ren H. Emerging important roles of circRNAs in human cancer and other diseases. Genes Dis 2021; 8: 412-23.
- 9. Aufiero S, Reckman YJ, Pinto YM, Creemers EE. Circular RNAs open a new chapter in cardiovascular biology. Nat Rev Cardiol 2019; 16: 503-14.
- Kristensen LS, Andersen MS, Stagsted LVW, Ebbesen KK, Hansen TB, Kjems J. The biogenesis, biology and characterization of circular RNAs. Nat Rev Genet 2019; 20: 675-91.
- 11. Ma H, Xu Y, Zhang R, Guo B, Zhang S, Zhang X. Differential expression study of circular RNAs in exosomes from serum and urine in patients with idiopathic membranous nephropathy. Arch Med Sci 2019; 15: 738-53.